

## Custom LNA Oligonucleotides



### For experiments requiring custom-designed, LNA-enhanced oligonucleotides

- Customize your own LNA oligonucleotides
- Use your own design or let QIAGEN's LNA experts design them for you
- Select from a wide range of labels and modifications
- Benefit from the easy-to-use online design tool

Custom LNA Oligonucleotides are ideal for studies involving short or very similar sequences. The high affinity of an [LNA-enhanced](#) oligonucleotide to its complementary sequence results in dramatically improved specificity and sensitivity, when compared with traditional DNA or RNA oligos. In many cases, LNA-enhanced oligonucleotides can be used to distinguish between sequences differing by only a single nucleotide, a feature that can be critical for the success of many experiments.

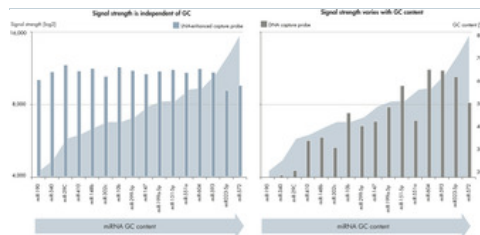
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### The power of $T_m$ normalization.

Signal intensity from microarray experiments using LNA-enhanced or DNA-based capture probes. miRNA targets with varying GC content were added at 100 amol each. The signal from DNA-based capture probes varies with GC content and results in poor detection of many miRNAs, whereas LNA probes offer robust detection of all miRNAs.



### Performance

LNA oligonucleotides exhibit unprecedented thermal stability when hybridized to a complementary DNA or RNA strand. For each incorporated LNA monomer, the melting temperature ( $T_m$ ) of the duplex increases by 2–8°C (see figure [Replace DNA with LNA for higher melting temperature](#)). In addition, LNA oligonucleotides can be made shorter than traditional DNA or RNA oligonucleotides and still retain a high  $T_m$ . This is important when the oligonucleotide is used to detect small or highly similar targets.

Since LNA oligonucleotides typically consist of a mixture of LNA and DNA or RNA, it is possible to optimize the sensitivity and specificity by varying the LNA content of the oligonucleotide. Incorporation of LNA into oligonucleotides has been shown to improve sensitivity and specificity for many hybridization-based technologies including PCR, microarrays and in situ hybridization (ISH).

$T_m$  normalization enables robust detection, regardless of GC content. The  $T_m$  of a nucleotide duplex can be controlled by varying the LNA content. This feature can be used to normalize the  $T_m$  across a population of short sequences with varying GC content. For AT-rich nucleotides, which give low melting temperatures, more LNA is incorporated into the LNA oligonucleotide to raise the  $T_m$  of the duplex. This enables the design of LNA oligonucleotides with a narrow  $T_m$  range, which is beneficial in many research applications such as microarrays, PCR and other applications in which sensitive and specific binding to many different targets must occur under the same conditions simultaneously. The power of  $T_m$  normalization is

## Principle

Use the guidelines below when designing your own Custom LNA Oligonucleotides:

LNA will bind very tightly to other LNA residues. Avoid self-complementarity and cross-hybridization to other LNA-containing oligonucleotides

Keep the GC content between 30–60%

Avoid stretches of more than 4 LNA bases, except when very short (9–10 nucleotides) oligonucleotides are designed

Avoid stretches of 3 or more Gs or Cs

For novel applications, design guidelines may have to be established empirically

## Applications

LNA oligonucleotides can be successfully used in a wide range of applications, including:

- miRNA research
- Small RNA research
- SNP genotyping
- mRNA antisense oligonucleotides
- Allele-specific PCR
- RNAi
- DNAzymes
- Fluorescence polarization probes
- Molecular beacons
- Microarray gene expression profiling
- Gene repair/exon skipping
- Splice variant detection
- Comparative genome hybridization (CGH)



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